

Exhibit 1

9/22 Sequencing gel of SEC2, T7, 1259C 3398

Transfection of DL plasmids

Plasmid	ml used
pCDNA1	10
pCDNA1 FT3m	10
pCDNA1 FT3m, F6, 3 (3+2)	10
pCDNA1 FT3m, F6, 4 (5+9)	4
pCDNA1 FT3m, F6L (I-1)	12
pCDNA1 FT6m F3, 4 (12+3)	5
pCDNA1 FT6m F3, 4, 5	20

Standard DEAE Dextran transfection protocol on COI-7 cells

DATA harvested for use in above Transfection	200	200	50	200
5+9	255	133	2.5	1.86
12+3	1237	143	2.4	1.65
12+7	1248	144	2.4	1.70

9/22 log gel of SEC-2 samples

SEP5 9925

SEP6 1259, T7, 3398B

Begin Sequencing FT7 clone - 10492/80RT

Primers	
8993	3244 (Not Work)
714	8904
946	8902
7931	8875
8661A	8851
8953	8771

9/23 Sequencing gel (Formamide vol 40%) of the above samples

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III TA cloning (Ampligen)

Followed instructions to letter

	μl	μl	μl	
10x	1.0	1.0		
10x	2.0	2.0		
10x	1.0			
10x	5.0	1.0		
10x	1.0	1.0	10 μg	
Transformant				thaw cells
10x	1.0	1.0	1.0	2 μl β-MEOL
Cells	50	50	50	Mix by TAP
Brill	450	450	450	Add DNA

Also transform KG's PCR products cloned into pCDHAT
- MCHD61/P3

Set up PCR experiment (in KG's Project)
use Vector primers; 3rd exon primers
to evaluate 14-7 FD DNA from library (PLS) pCD
2nd amp 1.1 μg/μl
this will tell if insert is in the library

Primers
5315 } pCDH7 Vector primers
5316
152
153

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II/1 Completed the assembly and checking of any
sequence discrepancies in FT7 Sequence
This is 3594 nt Sequence
from 300 → end in 2 directions
Save this sequence to SN. to check against his
independently determined sequence
II/2 no differences detected - full 99.9% confident
this sequence